- 1 The effect of milk on recovery from repeat-sprint cycling in female team-sport athletes
- 3 Paula Rankin^{1,5}, Michael J. Lawlor¹, Frank A. Hills², Phillip G. Bell³, Emma J.
- 4 Stevenson⁴ and Emma Cockburn^{5,6}
- ¹Department of Science and Health, Institute of Technology Carlow, Ireland
- ²Department of Natural Sciences, Middlesex University, UK
- 7 GlaxoSmithKline Human Performance Laboratory, UK
- 8 ⁴Institute of Cellular Medicine, Newcastle University, UK
- 9 ⁵London Sports Institute, Middlesex University, UK
- 10 ⁶School of Biomedical Sciences, Newcastle University, UK
- 12 Corresponding Author:
- 13 Paula Rankin

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- 14 Institute of Technology Carlow, Ireland
- Phone: 00 353 599175540. Email: paula.rankin@itcarlow.ie
- 16 Co-authors
- 17 Michael Lawlor (michael.lawlor@itcarlow.ie)
- 18 Frank Hills (f.Hills@mdx.ac.uk)
- 19 Phill Bell (phill Bell (phillip.x.bell@gsk.com)

- 20 Emma Stevenson (<u>emma.stevenson@newcastle.ac.uk</u>)
- 21 Emma Cockburn (emma.cockburn@newcastle.ac.uk)
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23 Abstract

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The consumption of milk post-eccentric exercise attenuates the effects of muscle damage in team-sport athletes. However, participation in team sport involves both concentric-eccentric loading and metabolic stress. Therefore, the aim of this study was to investigate the effects of post-exercise milk consumption on recovery from a cycling protocol designed to simulate the metabolic demands of team sport. Ten female team-sport athletes participated in a randomised cross-over investigation. Upon completion of the protocol participants consumed 500ml of milk (MILK) or 500ml of an energy-matched carbohydrate (CHO) drink. Muscle function (peak torque, rate of force development (RFD), countermovement jump (CMJ), 20m sprint), muscle soreness and tiredness, serum creatine kinase (CK), (high-sensitivity Creactive protein (hsCRP) and measures of oxidative stress (protein carbonyls (PC) and GSH:GSSG (oxidized glutathione:reduced glutathione) ratio) were determined pre-, 24h, 48h and 72h post-exercise. MILK had a possible beneficial effect in attenuating losses in peak torque (180°/s) from baseline to 24h (3.2±7.8% v -6.2±7.5%, MILK v CHO) and a possible beneficial effect in minimising soreness (baseline-48h; baseline-72h) and tiredness (baseline-24h; baseline-72h). There was no change in oxidative stress following the exercise protocol, though a *likely benefit* of milk was observed for GSH:GSSH ratio at baseline-24h (0.369 x/÷ 1.89, 1.103 x/÷ 3.96, MILK v CHO). MILK had an unclear effect on all other variables. Consumption of 500ml milk post-repeat sprint cycling had little to no benefit in minimising losses in peak torque, or minimising increases in soreness and tiredness and had no effect on serum markers of muscle damage and inflammation. Key words: MUSCLE DAMAGE, RECOVERY, PROTEIN METABOLISM, FEMALE ATHLETE, TEAM SPORT

Introduction

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Participation in exercise is known to result in both mechanical and metabolic stress. Mechanical stress, primarily consequential to eccentric muscle actions, results in morphological changes in the muscle (Lauritzen et al. 2009), with early damage occurring immediately following the muscle insult (Friden et al. 1983) and further damage evident 1-3 days after the exercise stress (Clarkson and Sayers 1999). Metabolic stess is evident by increases in metabolic flux through glycolytic and oxidative pathways, and increases in blood lactate, inflammation, oxidative stress, and markers of oxidative damage in the post-exercise period (Powers et al. 2011). Such stresses lead to a subsequent decline in muscle function and increases in muscle soreness which hinders the ability to perform exercise. In order to facilitate a faster recovery and maintain subsequent training volumes, intensity and performance, athletes employ a range of recovery strategies (Howatson and van Someren 2008). Nutritional interventions are one such strategy and there is evidence that the consumption of milk following eccentric exercise can attenuate decreases in muscle function in males (Cockburn et al. 2008, 2010, 2012) and females (Rankin et al. 2015) and limit increases in muscle soreness and creatine kinase, perhaps by enhancing protein synthesis post-exercise or limiting protein degradation. However, these investigations employed an eccentric exercise protocol on an isolated muscle group resulting in higher levels of mechanical stress and lower levels of metabolic stress than is normally observed following exercise participation (Silva et al. 2013). Metabolic stress leads to diverse physiological processes and may lead to different effects on muscle performance and markers of muscle damage. No studies have examined the effects of milk on recovery from exercise that primarily induces a metabolic rather than mechanical stress.

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Milk is a source of carbohydrate and anti-oxidants including Vitamin E, Vitamin A and glutathione (Haug et al. 2007) which are effective in reducing lipid peroxidation (Finaud et al. 2006). Moreover, milk is a source of whey protein that contains cysteine and methionine amino acids which are necessary for glutathione (GSH) production thus enhancing the antioxidant defence system (Madureira et al. 2007; Elia et al. 2006; Marshall 2004). Previous animal research concluded that supplementation with whey protein over a six week period prevented oxidative stress induced by heavy exercise (Elia et al. 2006). Still, these studies have examined long term supplementation with whey and not a single post-exercise intake nor its impact on muscle function or soreness In addition, unlike many other carbohydrate-protein recovery drinks, milk is an excellent source of casein. It has been suggested that milk's antioxidant capacity comes mainly from its casein fractions (Zulueta et al. 2009) due to the high amounts of antioxidant amino acids, including tyrosine, tryptophan, histidine, lysine and methionine present in casein (Rival et al. 2001). However, no studies have specifically examined casein's effects on oxidative stress following exercise. Some previous research has shown that carbohydrate or carbohydrate-protein mixtures do not reduce exercise-induced oxidative stress following endurance exercise (Karolkiewicz et al. 2001; McAnulty et al 2003; Goldfarb et al. 2009). However, McAnulty et al. (2007) reported lower oxidative stress levels when carbohydrate was consumed during both continuous cycling and cycling with intermittent rest periods. Furthermore, Kerasioti et al. (2012) reported that the consumption of a carbohydrate-protein cake immediately post-exercise (2h steady-state cycling) and at three further 1-hour intervals before a second exercise bout, reduced lipid peroxidation. The disparate results from these investigations may be because of different exercise modes and varied methods for the determination of oxidative stress and oxidative damage. Furthermore, these investigations focused on immediate recovery (1-24h),

and did not investigate the effect of the observed stress on subsequent muscle function and soreness, which is of significance to the training athlete.

Participation in a team sport results in muscle damage, inflammation and oxidative stress (Ascensão et al. 2008; Fatouros et al. 2010; Silva et al. 2013; Marin et al. 2011) and a subsequent decline in performance (Silva et al. 2013). As previous research has identified milk as being beneficial for recovery from isolated eccentric exercise, investigating the effect of milk on recovery from metabolic stress may provide insight in to the mechanisms by which milk might be beneficial for team sport athletes, and also offer an understanding of the relationships between metabolic stress, oxidative stress, inflammation and performance.

Therefore the aim of this study is to investigate the effect of milk on recovery from metabolic stress in team sport participants utilising a protocol designed to simulate the isolated metabolic demands of team sport. We hypothesise that post-exercise consumption of milk will attenuate decreases in muscle function and markers of muscle damage and oxidative stress.

Materials and Methods

Participants

Ten female team sport (camogie and ladies gaelic football) athletes (mean age $22.1 \pm 1.8 \text{ y}$) were recruited to take part in this study. Mean (\pm SD) height and mass was $162.7 \pm 9.0 \text{ cm}$ and $61.9 \pm 8.1 \text{ kg}$, respectively. All participants completed a medical health screening questionnaire and were excluded from the study if they met any of the following criteria: intolerance to dairy or lactose products, lower limb or back injury in the previous three months, surgery in the previous 6 months, known coronary disease, uncontrolled metabolic disorder or respiratory disease, pregnancy or post-partum. All participants were provided

with verbal and written briefings, following which written informed consent was recorded. Ethical Approval was provided by the London Sport Institute Ethics Sub-Committee at Middlesex University, and the Institute of Technology Carlow, where the data collection took place.

Study design

The study utilised a randomised cross-over design, with participants attending a familiarisation session plus two blocks of four days of testing. All participants completed the trials during two consecutive follicular phases of their menstrual cycle as in previous investigations utilising female participants (Dannecker et al. 2013; Rankin et al. 2015). All testing took place at the same time of day following an overnight fast and 24h abstinence from alcohol, caffeine and exercise. Briefly, the first laboratory visits comprised of familiarisation with initial measurements of muscle function (peak torque, rate of force development (RFD), countermovement jump height (CMJ), 20m sprint time), and familiarisation with an intermittent cycling protocol (standardised warm-up, five 2-min blocks of the protocol and one 15s maximal sprint) following which participants were randomly assigned to their group order.

Between 3 and 5 days later participants completed baseline measures of muscle function, soreness and provided a blood sample for analysis of muscle damage (CK), inflammation (hsCRP) and oxidative stress (Protein Carbonyls and GSG:GSSH ratio). This was followed by completion of an intermittent cycling protocol lasting approximately 60min designed to induce metabolic damage. Immediately upon completion of the protocol participants consumed 500ml of milk (MILK) or 500ml of an energy-matched carbohydrate drink (CHO) and were instructed not to consume any other fluid or food for a period of two hours. Participants returned to the laboratory to repeat the baseline measures at 2h (blood sample

only), 24h, 48h and 72h post-exercise. Participants were requested to refrain from any other strenuous activity for the duration of the study, and from treating the symptoms of muscle soreness and tiredness with interventions such as massage, cryotherapy, nutritional supplements and non-steroidal anti-inflammatory drugs.

Nutritional Intervention and Dietary Control

Immediately following the cycling protocol participants were provided with either 500ml of milk (Avonmore 1% Light, Glanbia, Kilkenny, Ireland) or 500ml of an energy-matched carbohydrate solution, which was consumed within 30min. Macronutrient composition (per 500ml) of the milk was as follows: Energy 910kJ/215kcal, Protein 17.0g, Carbohydrate 25.5g, Fat 5.0g. A volume of 500ml was chosen based on previous research (Cockburn et al., 2010, 2012). From an applied perspective, it was felt that 500ml was an easily consumed volume and that consumption of larger volumes may lead to stomach fullness and discomfort. The energy-matched carbohydrate solution consisted of glucose (52.6g) mixed with water and a commercially available orange flavoured fruit cordial (Nutritional information per 100ml of fruit cordial: Energy 37kJ/9kcal, Protein 0.2g, Carbohydrate 0.8g, Sodium Trace; MiWadi, Dublin, Ireland).

In order to maintain dietary control, participants completed a food diary for 24h prior to baseline testing and for the subsequent 3 days, and were asked to repeat the same diet for the second block of testing. Each participant was provided with a weighing scale and measuring jug for the duration of the study and was instructed to follow their usual eating habits before and during the investigation.

Metabolic stress protocol

Metabolic stress was induced by the completion of the Cycling Intermittent Sprint Protocol (CISP, Hayes et al. 2013) designed to simulate the metabolic demands of a repeat sprint sport. The protocol was modified slightly to include additional sprints (4x15s) which has been reported to result in oxidative stress (Jówko et al. 2014).

Thus the exercise protocol employed for the current study comprised of a standardised warm-up (5 min at 95 W and two 30-s bouts at 120 W with 30-s rest in between) followed by 2 x [14 x 2min bouts of exercise comprising of 10s of passive rest, 5s of maximal sprinting and 105s of active recovery, with a 15s maximal sprint followed by 1min active recovery taking place after the 7th and 14th 2min bout](Figure 1). The exercise bouts were separated by a 10min 'half-time' period during which the participants were permitted to dismount the ergometer and walk around in order to reduce venous pooling in the lower extremities, and minimise feelings of light-headedness and nausea. The total time for each 'half' was 30min and 30s, thereby simulating the ~30min halves of ladies gaelic football and camogie, sports from which the majority of participants were drawn. Power output, cadence, speed, HR and RPE were recorded throughout the exercise bout. Participants returned to complete the second cycling trial during the follicular phase of their next menstrual cycle, following which the drink not consumed on the first trial was ingested.

Blood sampling

On each day, prior to any other measures and following a 10min rest, a blood sample was collected by venepuncture from a forearm vein in ethylenediaminetetraacetic acid (EDTA), heparin and serum separator (SST) tubes. The samples were then centrifuged, aliquoted and stored at -80°C for later analysis of creatine kinase (CK), GSSG:GSH (oxidized glutathione:reduced glutathione) ratio, protein carbonyls (PC) and high sensitivity C-reactive protein (hsCRP). A blood sample was also collected 2h post completion of the exercise

protocol. Total serum CK and hsCRP were measured using high sensitivity procedures (Roche Cobas 6000 chemistry module c501, Hoffmann-La Roche, Basel, Switzerland). PC were determined using a commercially available kit (Abcam, Cambridge, UK) utilising a methodology based on that described by Levine et al (1990). Briefly samples were derivatised initially with dinitrophenylhydrazine (DNPH) producing functional groups into the oxidized protein which were detected by spectrophotemetry. The samples were washed to remove excess DNPH and the oxidised carbonyls were solubilised with guanidine prior to their detection by measuring absorbance at 375 nm. Finally, the protein content in each sample was measured so that the carbonyl content was expressed in terms of nmol/mg of total protein. The intra- and inter-assay variation for this kit is <7% and <10%, respectively. For the determination of GSH:GSSG using a commercially available kit (Abcam, Cambridge, UK) samples were first deproteinised by centrifugation through a 10kDa spin column. Thiol Green Indicator Reaction mix was then added which becomes strongly fluorescent upon reacting directly with glutathione. Samples were incubated prior to the measurement of fluorescence. Intra- and inter- assay CVs for this procedure are <15%.

Muscle Soreness and Muscle Tiredness

Active muscle soreness during squatting to approximately 90° knee flexion was measured on a visual analogue scale (VAS), with participants rating their level of soreness on a scale of 0 (no soreness) to 10 (as bad as it could be), as in previous research (Rankin et al. 2015). A similar VAS was used to measure muscle tiredness, with 0 indicating no tiredness, and 10 indicating as tired as could be.

210 Peak Torque

To determine peak torque (Nm), following a standardised warm-up, participants completed three dominant leg maximal effort knee extension repetitions at 60°/s and 180°/s, with 60s recovery between speeds on a Biodex System 3 Isokinetic dynamometer (Biodex Medical System, NY, USA). All participants were instructed to give maximal effort and to complete full range of motion at the knee for each repetition. Interclass correlations for this protocol at IT Carlow are 0.83-0.94.

Rate of force development

Rate of force development (RFD) was determined over the first 200ms of an isometric contraction, according to previous studies (Aagard et al. 2002). Briefly, two maximal 5s isometric contractions of the dominant leg quadriceps were performed on the same isokinetic dynamometer used for isokinetic peak torque measurements, with the knee fixed at an angle of 70° (0° = full extension). Participants were instructed to contract and 'push away as fast and forcefully as possible'. Any repetitions that showed a countermovement (a visible drop in the force signal) were excluded; otherwise RFD was determined from the repetition with the highest torque measurement. RFD was calculated over the time interval of 0-200ms (Δ torque/ Δ time) relative to the onset of contraction, which was defined as the time point when the torque generated exceeded the baseline by >7.5Nm (Aagard et al. 2002).

Countermovement Jump Height

Countermovement jump height was measured in cm using an Optojump optical measurement system (Microgate, Bolzano, Italy). Participants completed three trials employing standard countermovement jump technique, where they were instructed to flex their knees to approximately 90° and immediately jump for maximal height. Jump height was calculated from flight time and the highest recorded jump was used for analysis.

Sprint performance

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Twenty metre sprint performance, from a standing start 20 cm behind the start line, was recorded using timing gates (Microgate Racetime 2, Bolzano, Italy). Participants were instructed to sprint through the timing gates as fast as possible and completed three sprints with a rest time of 120s between sprints. Each participant's best time from three trials was used for analysis.

Data analysis

Data was analysed by making probabilistic magnitude based inferences about the true values of outcomes as described by Batterham and Hopkins (2006). By defining the smallest practical or biological effect, the probability of a worthwhile effect with inferential descriptions is possible. Within-group effects over time were determined using a published spreadsheet (Hopkins 2006) and the effect of MILK versus CHO was analysed using a second spreadsheet for analysis of a controlled trial (Hopkins 2006). Comparisons were made between baseline and 2h, 24h, 48h, and 72h post-exercise. Data for peak torque, countermovement jump and 20m sprint were log-transformed to overcome heteroscedastic error (Nevill and Lane 2007). Muscle soreness values were not log-transformed because of interval scaling (Nevill and Lane 2007). Means of log-transformed data were then back transformed to provide mean percentage change and percentage SD. Serum markers are however, reported as factors because of large percentage changes (Hopkins 2003). The smallest worthwhile effect was the smallest Cohen change in the mean: 0.2 times the between-subject SD for the baseline values of all participants (Batterham and Hopkins 2006). Chances of benefit and harm were assessed qualitatively as follows: <1% almost certainly none, 1-5% very unlikely, 5-25% unlikely, 25-75% possibly, 75-95% likely, 95-99% very likely, >99% almost certainly (Hopkins 2002).

Results

Preliminary Data

Analysis of participants' food diaries (Nutritics Professional Diet Analysis, Nutritics Ltd., Dublin, Ireland) indicated that there were no differences in energy, carbohydrate, protein or fat intake between trials (P>0.05). Analysis of the repeat-sprint exercise bouts revealed no significant differences (P>0.05) between trials on performance or physiological measures (Table 1).

Within-group effects

Analysis of within-group effects revealed post-exercise decreases in peak torque, CMJ, sprint performance and RFD, increases in serum CK and hsCRP and in muscle soreness and tiredness. No change was observed in PC or GSH:GSSG ratio. Mean effects \pm 90%CI, with qualitative inferences, are presented in Table 2.

Between-group effects

Peak torque

Baseline knee extension peak torque values of the dominant leg at 60° /s for the milk condition and the carbohydrate condition were 143.1 ± 25.5 Nm and 141.1 ± 23.9 Nm respectively. Percentage changes in peak torque at 60° /s for dominant knee extension can be seen in Figure 2. With a Smallest Worthwhile Effect (SWE) of 3.8%, changes in the peak torque of the dominant leg at 60° /s between baseline and 24h for the milk condition and carbohydrate condition were $-3.9 \pm 4.5\%$ and $-3.1 \pm 4.3\%$ respectively, a trivial effect for

- 279 CHO. Unclear outcomes for the comparison of MILK versus CHO were found at baseline-
- 280 48h (-6.4 \pm 12.2 % and -4.3 \pm 4.4 %) and baseline-72h (-4.4 \pm 9.5 % and -2.4 \pm 8.5 %).

- At baseline the peak torque values at 180°/s for the dominant leg of participants for MILK
- and CHO were 97.7 ± 16.6 Nm and 95.5 ± 16.0 Nm respectively. Changes in the peak torque
- of the dominant leg at 180°/s between baseline and 24h for MILK and CHO (SWE: 3.7%)
- were -3.2 ± 7.8 %, and -6.2 ± 7.5 % respectively, a possible benefit of milk. There was an
- unclear outcome for MILK (-5.0 \pm 12.7 %) versus CHO (-3.1 \pm 8.9 %) at baseline-48h. A
- possible negative effect was observed for MILK (-7.3 \pm 12.7 %) compared to CHO (-0.8 \pm
- 288 9.2%) at baseline-72h.
- 289 Rate of Force Development
- 290 Immediately prior to the repeat-sprint cycling exercise the mean rate of force development
- 291 was $433.1 \pm 159.1 \text{ Nm.s}^{-1}$ and $405.6 \pm 106.1 \text{ Nm.s}^{-1}$ for the MILK and CHO conditions
- 292 respectively. The SWE was 9.4% and unclear outcomes for MILK versus CHO were found at
- baseline-24h (-5.5 \pm 25.3 % v -10.5 \pm 22.0 %), baseline-48h (-1.9 \pm 16.4 % v -2.7 \pm 20.3 %)
- and baseline-72h ($-0.5 \pm 30.7 \% \text{ v } 6.3 \pm 28.7\%$).
- 295 *20m sprint*
- Baseline 20m sprint times for the milk condition and carbohydrate condition were 3.58 ± 0.12
- s and 3.59 ± 0.11 s respectively and the SWE was determined as 0.7%. Unclear outcomes
- were found for MILK versus CHO at baseline-24h (1.8 \pm 2.2 % and 1.0 \pm 1.3 %), baseline-
- 299 48h $(1.0 \pm 2.8 \%)$ and $0.6 \pm 2.5 \%$ and baseline-72h $(0.2 \pm 2.2 \%)$ and $-0.4 \pm 1.2 \%$).
- 300 Countermovement jump performance

Immediately prior to the cycling exercise the mean countermovement jump heights of the milk and carbohydrate conditions were 28.9 ± 2.9 cm and 28.8 ± 2.4 cm respectively, with a SWE of 1.9%. Unclear outcomes for MILK versus CHO were found at all time points, baseline-24h (-1.6 \pm 4.7 % v -2.0 \pm 3.1 %), baseline-48h (-1.4 \pm 4.6 % and -1.9 \pm 5.4 %) and baseline-72h (-0.2 \pm 4.1 % v-0.0 \pm 4.7 %). A summary of the statistical analysis for Peak Torque, RFD (0-200ms) of the dominant leg, 20m sprint performance and countermovement jump performance can be seen in Table 3.

Soreness

- The exercise protocol resulted in a small increase in muscle soreness for both conditions, peaking at 24h for MILK and 48h for CHO (Figure 3). By 72h soreness had almost returned to zero. A comparison of changes in soreness for the period baseline-24h indicated an unclear outcome for MILK (7.0 \pm 8.2 %) compared to CHO (12.0 \pm 15.5 %). A possible benefit of MILK versus CHO was seen at baseline-48h (4.0 \pm 7.0 % v 14.0 \pm 21.7%) and baseline-72h (1.0 \pm 3.2 % v 8.0 \pm 10.2 %)
- 316 Tiredness

Muscle tiredness increased over time for both conditions but by 72h tiredness had almost returned to zero. A comparison of changes in muscle tiredness from baseline to 24h indicated a possible benefit for the consumption of MILK (18.0 \pm 11.4 %) compared to the consumption of CHO (24.0 \pm 15.8 %). An unclear outcome was observed from baseline to 48h (16.0 \pm 12.6 % versus 24.0 \pm 20.7 % for MILK and CHO conditions respectively). Changes in tiredness from baseline-72h showed a possible benefit of MILK (1.0 \pm 3.2 %)

- compared to CHO (9.0 \pm 13.7 %). A summary of the statistical analysis for muscle soreness
- and muscle tiredness can be seen in Table 3.
- 325 Creatine Kinase
- Serum CK values prior to exercise were 155.0 ± 75.4 U/l and 138.0 ± 62.6 U/l for the milk
- and carbohydrate conditions respectively. Unclear outcomes for the comparison of MILK
- 328 versus CHO were found at baseline-2h (1.52 x/ \div 1.27 v 1.41 x/ \div 1.17), baseline-24h (1.30
- 329 $x/\div 1.55 \text{ v } 1.26 \text{ x}/\div 1.65$), baseline-48h(1.03 $x/\div 1.57 \text{ v } 1.14 \text{ x}/\div 1.57$) and baseline-72h (0.99
- 330 $x/\div 1.56$ and $0.97 x/\div 1.76$).
- 331 *hsCRP*
- Baseline hsCRP values immediately prior to exercise were 1.16 ± 0.91 mg/l and 1.38 ± 0.99
- mg/l for the milk and carbohydrate conditions respectively. Unclear outcomes were observed
- 334 for MILK versus CHO at baseline-2h(1.00 x/÷ 1.07 v 1.02 x/÷ 1.17), baseline-24h (2.09 x/÷
- 335 1.75 v 1.42 x/ \div 2.03), baseline-48h (1.59 x/ \div 1.50 v 1.15 x/ \div 1.98) and baseline-72h (1.25
- 336 $x/\div 1.73 \text{ v } 0.925 \text{ x/}\div 1.99$).
- 337 Protein Carbonyls (PC)
- Protein Carbonyl values prior to exercise were 0.54 ±0.14 nmol/mg/protein for the MILK
- group and 0.49 ± 0.36 nmol/mg/protein for the CHO group. Unclear outcomes were observed
- 340 for MILK versus CHO; baseline-2h (1.10 x/ \div 1.54 v 0.88 x/ \div 2.36), baseline-24h (1.23 x/ \div
- 341 1.82 v 0.85 x/ \div 2.72), baseline-48h (1.33 x/ \div 1.23 v 0.76 x/ \div 1.99) and baseline-72h (0.88 x/ \div
- 342 1.46 and 0.957 $x/\div 3.04$).
- 343 GSH:GSSH ratio

Prior to the cycling exercise the mean GSH:GSSG ratio of the milk and carbohydrate conditions were 0.57 ± 0.27 and 0.98 ± 0.92 respectively. An unclear outcome for MILK versus CHO was found at baseline-2h (0.871 x/÷ 1.99 v 1.014 x/÷ 8.52). A likely benefit of MILK was observed at baseline-2h (0.369 x/÷ 1.89 v 1.103 x/÷ 3.96). Unclear outcomes were found at other timepoints: baseline-48h (0.827 x/÷ 3.15 and 1.105 x/÷ 8.800) and baseline-72h (0.751 x/÷ 6.39 v 0.686 x/÷ 7.72). A summary of the statistical analysis for Creatine Kinase, hs-CRP, PC and GSH:GSSG ratio can be seen in Table 3.

Discussion

Repeat-sprint cycling resulted in decrements in muscle function and increases in perceptions of soreness and tiredness, and serum markers of muscle damage and inflammation. However, no increase in oxidative stress was observed. The results indicate that the consumption of 500ml of milk post-intermittent sprint cycling exercise had minimal effect on recovery of muscle function, inflammation and markers of muscle damage. While a possible benefit of milk was observed for peak torque at 180°/s at 24h post-exercise, no effect was seen on any other muscle function variable. A possible negative effect for MILK was noted at 72h post-exercise for peak torque at 180°/s. The consumption of milk attenuated increases in muscle soreness and tiredness over 72h post-exercise, compared to the consumption of a volume and energy matched carbohydrate drink. Markers of oxidative stress did not increase following the exercise protocol.

A possible benefit for MILK was observed for Peak Torque at 180°/s at 24h post exercise, but at no other time and for no other muscle function variable, CK or hsCRP. The lack of effect of milk on muscle function, inflammation and CK is likely because of the nature of the exercise protocol. Previous research has found that the consumption of milk following eccentrically loaded exercise had a beneficial effect in attenuating decreases in peak torque,

sprint performance and reactive strength index (Cockburn et al. 2008, 2010, 2012; Rankin et al. 2015). Cycling employs concentric muscle actions (Bijker et al. 2002), resulting in metabolic rather than mechanical stress, and may explain the low level of muscle damage observed in this study. This lower level of damage is reflected in the smaller decreases in muscle performance observed across all variables (3.98%) compared to previous studies involving eccentric exercise with female team-sport athletes (6.95%, Rankin et al. 2015). Serum CK levels were elevated during the recovery period, peaking 24h post-exercise, and returning to baseline values by 72h with no difference between interventions. Mean peak CK values (225.6 \pm 72.3 and 193.6 \pm 61.0 IU/l for MILK and CHO respectively) were lower than observed for female athletes in investigations employing eccentric exercise (8873.0 \pm 13306.0 and 11697.7 \pm 8423.3 IU/l, MILK and CHO respectively, Rankin et al. 2015), though similar to other cycling studies investigating recovery interventions (Bell et al. 2014; Jowko et al. 2014). In summary, the observed levels of muscle damage were low, losses in muscle function were small and milk had no effect on minimising these effects.

It is possible that an increase in protein synthesis or a decrease in protein breakdown following the mechanical stress of eccentric loading in previous studies is enhanced with the intake of milk post-exercise, a mechanism not observed following isolated metabolic stress in this study. It is conceivable that the nature of the exercise (cycling) may have stimulated mitochondrial protein synthesis rather than myofibrillar protein synthesis. Wilkinson et al. (2008) noted a 67% increase in myofibrillar FSR following a resistance exercise session but no change following an endurance (cycling) session while Dumke et al (2009) reported stimulation of genes associated with mitochondrial biogenesis following cycling, an effect reported by others following high intensity sprint cycling protocols and training (Little et al, 2010; Granata et al., 2016; MacInnis et al, 2017). In the current study the measurements of muscle function over the recovery period (peak torque, RFD, CMJ, sprint performance) were

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not dependent on mitochondrial function and therefore any benefit of milk intake may not have been apparent from the measures chosen. It is possible that a larger intake of milk may have produced different outcomes. The amount of protein provided by the 500ml of milk was 17g which provided a mean relative intake of 0.27 g/kg, though given the range in body mass, the range in relative intake was 0.23-0.32 g/kg. The lower relative intake of protein may have been insufficient to maximally stimulate protein synthesis and attenuate muscle function losses. Nevertheless, Cockburn et al (2012) reported that 500ml of milk consumed post-exercise was as effective as 1000ml in attenuating the negative effects of EIMD. Further research is warranted to extrapolate this further.

Exercise causes a systemic inflammatory response with increases in cytokine and leucocyte activity (Febbraio and Pedersen 2005). hsCRP is an acute phase protein and its synthesis in the liver is triggered as an inflammatory response following increase cytokine secretion, most notably IL-6. Mean absolute hsCRP values were very similar in both trials, suggesting similar inflammatory responses. Not surprisingly, given that cycling is concentric in nature, the hsCRP response to the exercise protocol was considerably lower than that observed following resistance exercise (Bowtell et al. 2011) and marathon running (Clifford et al. 2016), suggesting that the magnitude of the inflammatory response is significantly influenced by the mode of exercise. The values, however, are comparable to that reported previously in cycling research (Roengrit et al. 2014) with peak hsCRP observed at 24h post-exercise, returning towards baseline values by 72h post-exercise. The magnitude of change was not different between the conditions, indicating that post-exercise consumption of milk did not modulate the inflammatory response after metabolic exercise. There is considerable disagreement in the literature regarding the effects of carbohydrate and carbohydrate-protein intake on inflammatory measures. For example, while Kerasioti et al. (2013) observed a decrease in inflammatory markers (IL-6, CRP) following the ingestion of a carbohydrate-protein cake,

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Cosio-Lima et al. (2012) reported no differences in inflammatory responses (TNF-α. IL-6) following the ingestion of carbohydrate versus carbohydrate-protein solutions. Such contradictory results are probably reflective of the variation in exercise protocols employed, the inflammatory markers measured or the specific composition of the nutritional interventions. It is important to note that inhibition of post-exercise inflammation is not always an aim of post-exercise nutritional intervention, as prevention of exercise-induced inflammation may inhibit muscular adaptation to exercise (Mastaloudis et al, 2004).

Interestingly, even though post-exercise muscle soreness and tiredness ratings were low compared to those following eccentric exercise, post-exercise milk consumption had a positive effect in attenuating increases in muscle soreness and tiredness over the 72h postexercise period. This is somewhat unexpected given that there were no differences in muscle function or inflammation levels between the groups. Inflammation is a proposed mechanism for the manifestation of muscle soreness (MacIntyre et al. 1995). Increased histamines and bradykinins sensitise nociceptors and increase the sensation of pain (Malm 2001). Oedema may also contribute to the sensation of soreness, with swelling exerting increased osmotic pressure within the fibres and further sensitising nociceptors (Malm 2001; Clarkson and Hubal 2002). The disparity of results observed in this study may indicate different pathways for the recovery of muscle function and perception of soreness and tiredness which may or may not involve the inflammatory response. It is possible that hsCRP may not be reflective of the total cytokine activity in the post-exercise period and that measurement of other cytokines such as TNF-α or IL-6 may have provided greater insight. Nonetheless, because this was not a blind study it is plausible that information bias may have occurred through knowledge of the nature of the allocated intervention (Booth et al. 1992), where participants may have been aware of previously published research highlighting potential benefits of milk for recovery, thus leading to a perception of attenuated soreness and tiredness.

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Surprisingly, the intermittent sprint protocol utilised in this study did not result in an increase in oxidative stress as indicated by PC and GSH:GSSG ratio, despite the considerable physical demand imposed by the exercise. This is in contrast with previous research that reported increases in post-exercise oxidative stress following sprint cycling (Jowko et al. 2014), though in agreement with Bloomer et al. (2007) and Farney et al. (2012). There are a number of possible reasons for this observation. Firstly, only selective biomarkers of oxidative stress were measured; the inclusion of additional markers or alternative assay techniques may have indicated an increase in oxidative stress. Both PC and GSH:GSSG have been identified as useful indicators of protein oxidation and redox disturbance respectively (Powers et al. 2010). Protein Carbonyls are produced when ROS attack amino acids and thus levels increase when oxidative stress levels increase. GSH is an antioxidant used in the reduction of lipid hydroperoxides generating oxidised glutathione (GSSG). When cells are exposed to oxidative stress levels of GSSG increase and the ratio of GSSG to GSH increases. Nevertheless the validity of many measures of oxidative stress has been questioned, with poor correlations between different markers of oxidative stress and between different methods for measuring individual markers (Finkler et al. 2014). Secondly, oxidative stress may have occurred in muscle tissue but was not detected in the serum samples as has been reported in animal research (You et al. 2005). Thirdly, a lack of observed oxidative stress may be because the participants were trained team-sport players, well-accustomed to the metabolic demands of repeat-sprinting all be it with a different exercise mode. This training may have resulted in adaptations that resulted in minimal change in PC and GSH:GSSG ratio over the duration of this study. Lending support to this idea Bloomer et al. (2006, 2012) reported no increase in oxidative stress in trained individuals following sprint cycling, and Bogdanis et al. (2013) reported attenuated oxidative stress responses following high intensity interval training. Finally, it is possible that a lack of oxidative stress may be because of the gender of the

participants. Previous research has indicated that females experience lower levels of oxidative stress than males (Ide et al. 2002) and that following exercise markers of oxidative stress return to baseline quicker than males (Mastaloudis et al. 2004). This lower level of oxidative stress may be because of smaller muscle mass, leading to lower levels of mitochondrial flux and lower ROS production (Ide et al. 2002). Additionally, Tiidus (1995, 2003) proposed that estrogen may act as an antioxidant against lipid peroxidation of the cell membrane during exercise. Estrogens possess a hydroxyl group on their A (phenolic) ring in a similar structure to tocopherol (Vitamin E) which is a known antioxidant (Ayres et al. 1998; Persky et al. 2000). Acting in a similar way to tocopherol, estrogen may donate hydrogen atoms leading to a termination of peroxidation chain reactions (Tiidus 1995; Ayres et al. 1998; Persky et al. 2000). Future investigations examining the effect of milk on oxidative stress should carefully consider exercise protocol design, training status and gender of the participants.

The majority of investigations examining the effects of nutritional interventions on recovery from metabolic stress employed a protocol that required participants to consume the nutritional substance over a prolonged period of time. For example, Samaras et al. (2014) observed an increase in glutathione levels when athletes were supplemented with a protein-carbohydrate bar for two months. The only investigation examining the effects of post-exercise consumption of a protein-carbohydrate supplement on oxidative stress markers was Kerasioti et al. (2012). While they observed a reduction in Thiobarbituric Acid Reactive Substances (TBARS) with the intake of an experimental cake post-2hours of cycling, there was no effect on subsequent time-trial performance. It is thus likely that the timing of antioxidant supplementation is an important factor that could influence exercise-induced oxidative stress (Goldfarb et al. 2009), and that post exercise intake of antioxidants does not have a positive effect on oxidative stress markers and may even result in a pro-oxidant response (Nieman et al. 2002). Thus it is unlikely that one-off post-exercise consumption of

nutritional products, as in the current study, will have any effect on recovery from oxidative stress. In this study the beverage was consumed only after the exercise bout, as this is a regular practice of team sport athletes.

In conclusion, the consumption of 500ml of milk post non-eccentric exercise had minimal effect on the attenuation of losses in muscle function and no effect on increases in markers of muscle damage and inflammation following repeat-sprint cycling, though consumption of milk did reduce perceptions of soreness and tiredness. Speculatively, the benefit of milk for athletic recovery is likely to be greatest following activities that have an eccentric component. Further research is warranted in this area to investigate the effects of milk on recovery from exercise that induces high levels of oxidative stress. However, recently it has been suggested that minimising oxidative stress may inhibit adaptation to exercise (Paulsen et al. 2014; Buresh and Berg 2015; Pingitore et al. 2015). From this perspective future research examining the effect of nutritional interventions on oxidative stress should carefully consider the practical application of outcomes. Consideration could be given to examining prolonged intake of milk prior to exercise performance and during the recovery period and the effect of milk on recovery from simulated or actual sport performance.

Acknowledgements and Conflict of Interest

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Table 1. Comparison of performance and physiological variables for cycling trials prior to Milk and Carbohydrate consumption

Variable	1 st half		2 nd half		Total	
variable	MILK	СНО	MILK	СНО	MILK	СНО
Avorago	117.1±	112.0±	109.3±	105.3±	113.2±	108.7±
Average						
power (W)	15.7	14.4	14.8	16.1	14.8	14.6
Peak power	695.4±	661.3±	638.6±	636.0±	667.0±	648.7±
(W)	135.6	124.1	135.4	129.4	129.4	130.3
Power/mass	2.0 ± 0.3	1.8 ± 0.2	1.8 ± 0.2	1.7 ± 0.2	1.9 ± 0.2	1.8 ± 0.2
(W/kg)						
Estimated	333.3±	$325.5 \pm$	$307.7 \pm$	$308.9 \pm$	320.5±	317.2±
EE (kcal)	24.2	25.2	27.6	37.3	24.4	26.7
Average	70.1 ± 3.9	67.7 ± 4.7	68.6 ± 4.2	66.5 ± 4.5	69.4 ± 4.0	67.1 ± 4.1
cadence						
(rpm)						
Peak	120.1±	117.8±	118.7±	115.0±	119.4±	116.4±
cadence	12.3	9.1	13.0	9.8	11.3	8.8
(rpm)						
Rev count	2255.2±	2207.8±	2128.0±	2121.7±	2191.6±	2164.8±
	124.1	128.6	136.6	215.7	112.4	130.6
Average	27.7 ± 1.5	27.3 ± 1.4	27.2 ± 1.5	26.6 ± 1.7	27.5 ± 1.4	27.0 ± 1.4
speed						
(kmh)						
Distance	14.5 ± 0.5	14.5 ± 1.1	14.0 ± 0.8	13.9 ± 0.8	14.2 ± 0.6	14.1 ± 0.9
covered						
(km)						
Average	1:58±	2:01±	2:00±	2:02±	2:00±	2:01±
pace	0:06	0:07	0:05	0:08	0:06	0:06
(min:ss/km)						
HR (bpm)	171.7±	169.9±	168.9±	167.5±	170.3±	168.7 ± 6.5
(1 /	10.9	8.2	9.7	5.8	10.1	
RPE	11.8 ± 1.6	11.9 ± 2.1	13.8 ± 1.8	14.1 ± 2.8	12.8 ± 1.4	13.0 ± 2.3

Table 2. Within-group effects over time for dependent variables

Variable Timeframe		Mean effect ± 90% CI	Qualitative inference
Peak Torque 60°/s			
Extension			
MILK	B-24	-4.0 ± 2.7	Possibly lower
	B-48	-7.1 ± 7.4	Likely lower
	B-72	-4.9 ± 5.6	Possibly lower
СНО	B-24	-3.2 ± 2.5	Possibly lower
	B-48	-4.4 ± 2.5	Possibly lower
	B-72	-2.7 ± 4.7	Possibly lower
Peak Torque 180°/s			
Extension			
MILK	B-24	-3.4 ± 4.5	Possibly lower
	B-48	-5.8 ± 8.1	Possibly lower
	B-72	-2.5 ± 1.7	Unclear
СНО	B-24	-6.5 ± 4.4	Possibly lower
	B-48	-3.5 ± 5.1	Likely lower
	B-72	-1.1 ± 5.3	Unclear
CMJ			
MILK	B-24	-1.7 ± 2.7	Possibly lower
	B-48	-1.5 ± 2.6	Possibly lower
	B-72	0.2 ± 2.4	Unclear
СНО	B-24	-2.0 ± 1.8	Possibly lower
	B-48	-2.0 ± 3.2	Possibly lower
	B-72	0.1 ± 2.8	Unclear
20m sprint			
MILK	B-24	-1.8 ± 1.2	Likely lower
	B-48	-0.9 ± 1.6	Possibly lower
	B-72	0.2 ± 1.3	Unclear
СНО	B-24	-1.0 ± 0.7	Likely lower
	B-48	-0.5 ± 1.5	Unclear
	B-72	-0.4 ± 0.7	Unclear
RFD			
MILK	B-24	-8.5 ± 14.6	Possibly lower
	B-48	0.8 ± 8.9	Likely trivial
	B-72	-3.9 ± 18.1	Unclear
СНО	B-24	-12.7 ± 11.9	Likely lower
	B-48	-4.9 ± 12.9	Unclear
	B-72	3.3 ± 15.0	Unclear
Creatine Kinase			
MILK	B-24	1.376 x/÷ 1.283	Likely increase
	B-48	1.165 x/÷ 1.401	Unclear
	B-72	1.096 x/÷ 1.323	Unclear
СНО	B-24	1.461 x/÷ 1.273	Very likely increase
	B-48	1.283 x/÷ 1.274	Likely increase
	B-72	1.083 x/÷ 1.508	Unclear

hsCRP			
MILK	B-24	2.163 x/÷ 1.353	Most likely increase
	B-48	1.685 x/÷ 1.259	Mostlylikely increase
	B-72	1.487 x/÷ 1.358	Likely increase
СНО	B-24	1.735 x/÷ 1.375	Very likely increase
	B-48	1.437 x/÷ 1.405	Likely increase
	B-72	1.087 x/÷ 1.582	Unclear
Soreness			
MILK	B-24	0.7 ± 0.5	Likely increase
	B-48	0.3 ± 0.4	Likely trivial
	B-72	0.1 ± 0.2	Most likely trivial
СНО	B-24	1.2 ± 0.9	Likely increase
	B-48	1.3 ± 1.3	Likely increase
	B-72	0.8 ± 0.9	Possible increase
Tiredness			
MILK	B-24	1.8 ± 0.7	Most likely increase
	B-48	1.5 ± 0.8	Very likely increase
	B-72	0.1 ± 0.2	Most likely trivial
СНО	B-24	2.4 ± 0.9	Most likely increase
	B-48	2.3 ± 1.3	Very likely increase
	B-72	0.9 ± 0.8	Likely increase
PC			
MILK	B-24	1.111 x/÷ 1.449	Unclear
	B-48	1.081 x/÷ 1.296	Unclear
		0.843 x/÷ 1.269	Unclear
СНО	B-24	0.940 x/÷ 1.925	Unclear
	B-48	0.761 x/÷ 1.929	Unclear
	B-72	0.716 x/÷ 1.727	Unclear
GSH: GSSG			
MILK	B-24	0.370 x/÷ 1.532	Unclear
	B-48	$0.750 \text{ x/} \div 2.381$	Unclear
	B-72	0.650 x/÷ 3.394	Unclear
СНО	B-24	1.356 x/÷ 3.910	Unclear
	B-48	1.105 x/÷ 7.949	Unclear
	B-72	$0.657 \text{ x/} \div 5.040$	Unclear

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771 Table 3. Effects on Peak Torque, RFD, sprint performance, CMJ, soreness, tiredness, CK,

hsCRP, PC and GSH:GSSG ratio following repeat sprint cycling exercise

Variable	Time Frame	Mean effect ^a ± 90% CI ^b	Qualitative Inference	
Peak Torque 60°/s	B – 24h	-0.3 ± 3.2	Likely trivial	
Dominant leg	B-24h B $-48h$	-0.3 ± 3.2 -2.4 ± 8.0	Unclear	
Dominant icg	B - 72h	2.0 ± 6.2	Unclear	
Peak Torque 180°/s	$\frac{B - 72H}{B - 24h}$	3.3 ± 6.5	Possibly beneficial	
Dominant leg	B - 2411 B $- 48h$	-2.5 ± 9.5	Unclear	
Dominant leg	B – 4811 B – 72h			
DED (0.200mg)		-7.1 ± 8.9	Possibly negative Unclear	
RFD (0-200ms)	B – 24h	2.0 ± 6.2		
Dominant leg	B – 48h	6.3 ± 16.4	Unclear	
20 0 : .	B – 72h	-4.4 ± 21.0	Unclear	
20m Sprint	B-24h	0.8 ± 1.4	Unclear	
performance	B-48h	0.4 ± 2.1	Unclear	
	B – 72h	0.6 ± 4.0	Unclear	
Countermovement	B-24h	0.3 ± 3.2	Unclear	
jump performance	B-48h	0.5 ± 4.0	Unclear	
	B-72h	-0.1 ± 3.5	Unclear	
Muscle soreness	B-24h	-0.4 ± 1.0	Unclear	
	B-48h	-1.0 ± 1.3	Possibly beneficial	
	B-72h	-0.7 ± 1.0	Possibly beneficial	
Muscle Tiredness	B-24h	-0.6 ± 1.1	Possibly beneficial	
	B-48h	-0.8 ± 1.3	Unclear	
	B-72h	-0.8 ± 0.8	Possibly beneficial	
CK	B – 2h	1.075 x/÷ 1.185	Unclear	
	B-24h	1.025 x/÷ 1.516	Unclear	
	B-48h	0.906 x/÷ 1.495	Unclear	
	B-72h	1.013 x/÷ 1.591	Unclear	
hs-CRP	B – 2h	0.983 x/÷ 1.111	Unclear	
	B-24h	1.475 x/÷ 1.720	Unclear	
	B-48h	1.377 x/÷ 1.593	Unclear	
	B-72h	1.347 x/÷ 1.696	Unclear	
PC	B – 2h	1.241 x/÷ 1.783	Unclear	
	B-24h	1.445 x/÷ 2.087	Unclear	
	B-48h	1.749 x/÷ 1.984	Unclear	
	B-72h	0.920 x/÷ 2.157	Unclear	
GSH:GSSG ratio	B – 2h	0.859 x/÷ 4.433	Unclear	
	B-24h	$0.334 \text{ x/} \div 3.228$	Likely Positive	
	B-48h	$0.749 \text{ x/} \div 8.474$	Unclear	
	B – 72h	$1.095 \text{ x/} \div 5.337$	Unclear	
O1:4-4: If		lihood that the true value		

Qualitative Inference represents the likelihood that the true value will have the observed magnitude

^a Mean effect refers to MILK minus CHO

 $^{^{\}rm b}$ ± 90% CI: add and subtract this number to the mean effect to obtain the 90% confidence intervals for the true difference

7	779	Figure Captions
7	780	Figure 1. Schematic of the Exercise Protocol
7	781	Figure 2. Peak torque at 60°/s for dominant knee extension in response to repeat sprint
7	782	cycling exercise for MILK (n=10) and CHO (n=10). Values are presented as means \pm SD.
7	783	Figure 3. Muscle soreness in response to repeat sprint cycling exercise for MILK (n=10)
7	784	and CHO (n=10). Values are presented as means \pm SD.
7	785	
7	786	
7	787	
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7	789	
7	790	
-	791	
7	792	
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7	794	

Figure 1

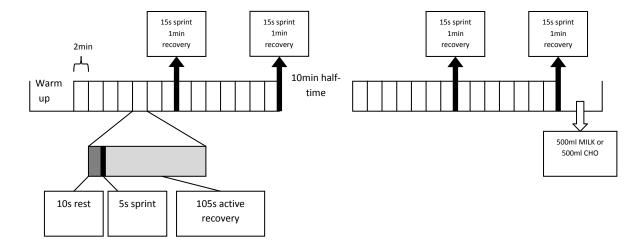


Figure 2

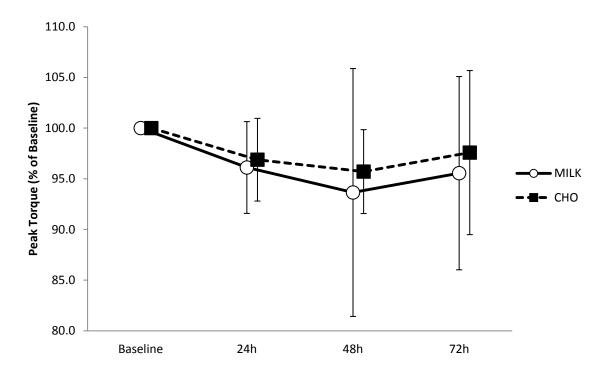


Figure 3

